

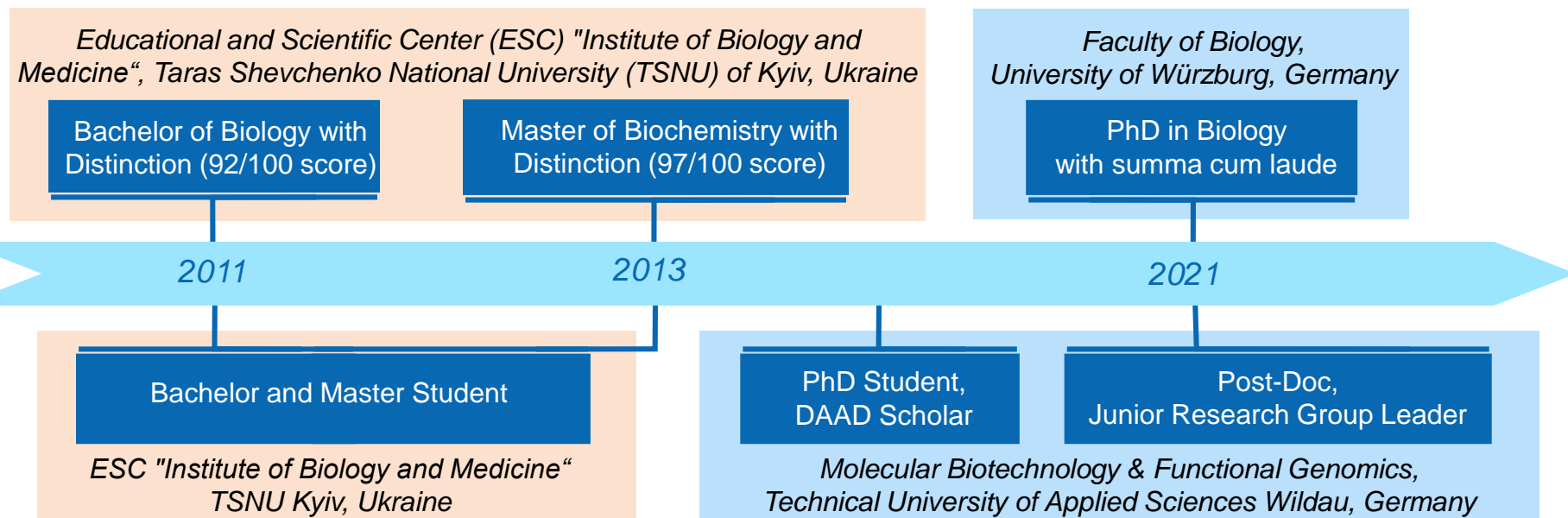
# Planning 1<sup>st</sup> experiments on chemical and biological effects of PITZ beam

for PITZ run during KW 44-45 (4-11.11.2022)

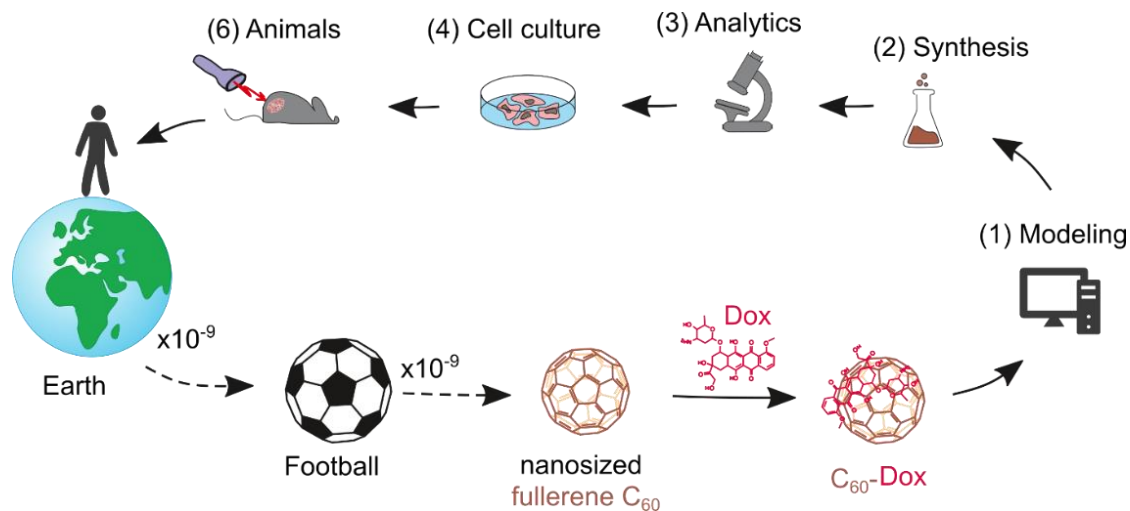
Anna Grebinyk  
Zeuthen, 20.10.2022



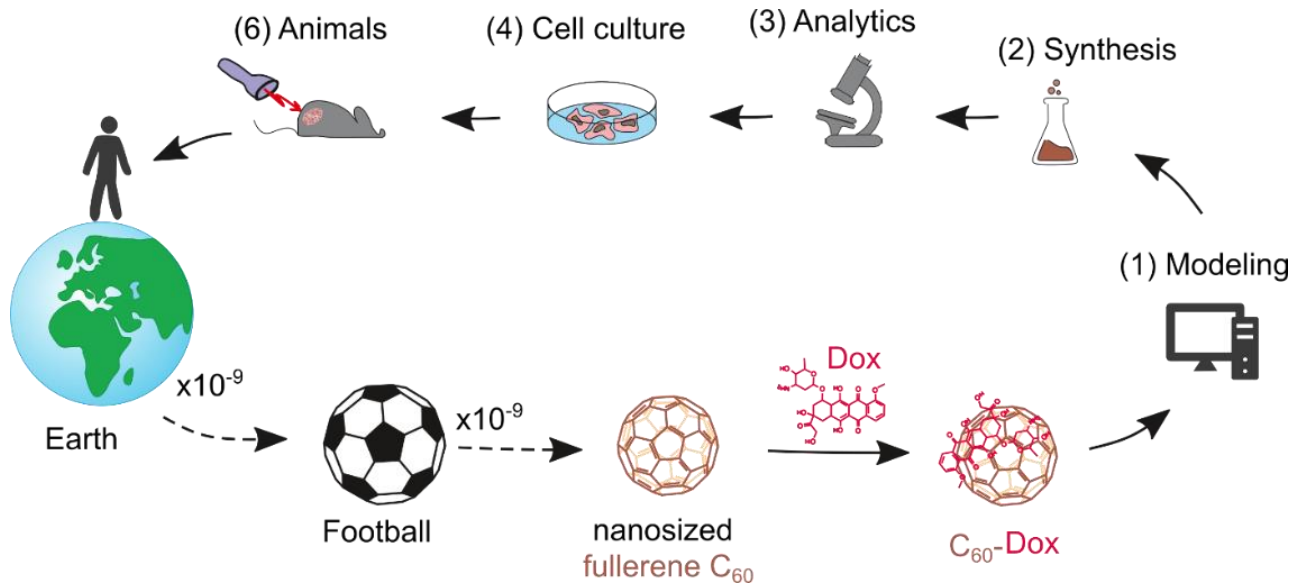
## Education & research experience



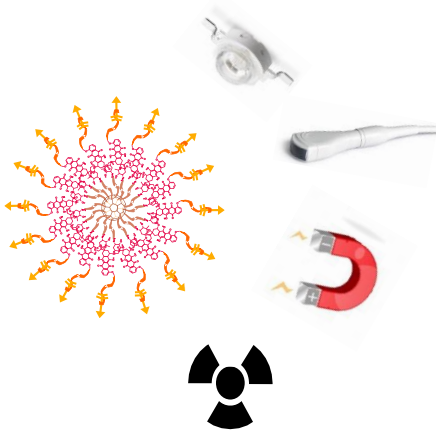
## Intradisciplinary background



# Intradisciplinary background

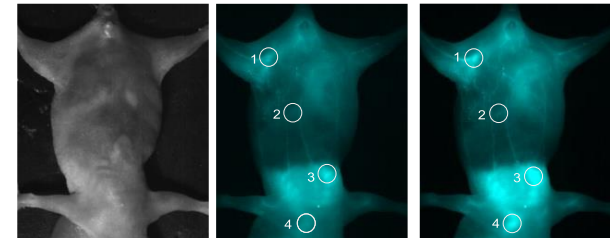


# Multimodal synergistic cancer therapy



- Chemotherapy
- Photodynamic therapy
- Photothermal therapy
- Sonodynamic therapy
- Magnetic hyperthermia
- Radiation therapy

+ Imaging



# R&D for Cancer Radiation Therapy (RT)

## Conventional RT

<5 Gy/min



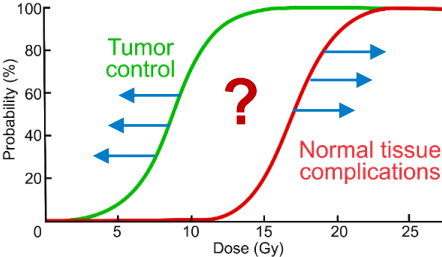
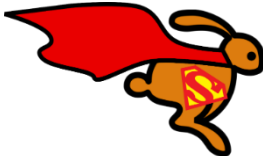
## FLASH RT

40-10<sup>9</sup> Gy/s



## FLASHlab@PITZ

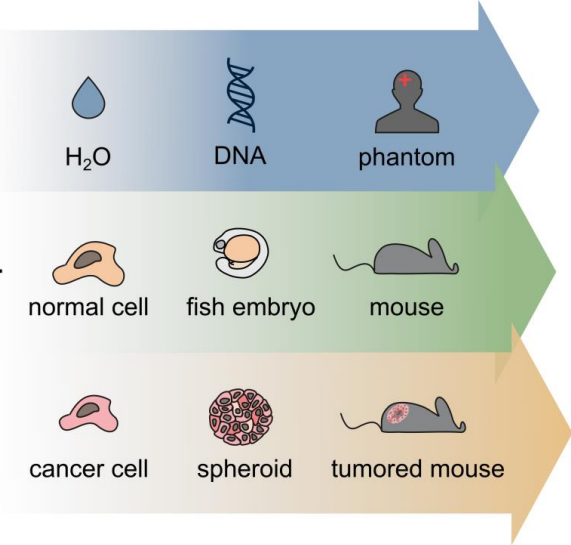
≤10<sup>14</sup> Gy/s



### Radiation



### Models



### Effects



# R&D for Cancer Radiation Therapy (RT)

Radiation

Models

Effects



H<sub>2</sub>O



DNA



phantom



normal cell



fish embryo



mouse



cancer cell



spheroid



tumored mouse



*in silico*  
*in vitro*  
*in vivo*

On behalf of the Organising Committee, we are delighted to confirm that your abstract has been selected for an **E-Poster Viewing**.

**Abstract Number: 443**

**Abstract Title: CHEMICAL EFFECTS OF FLASHLAB@PITZ BEAM**



# Chemical effects of PITZ beam

## H<sub>2</sub>O<sub>2</sub> production measurements during water radiolysis

- Amplex® Red Hydrogen Peroxide/Peroxidase Assay Kit (Invitrogen)
- One pretest measuring for checking timing



50/100 µL in  
0.5 mL tube

h: 7/10 mm

### Dose:

0, 25, 50, 75, 100 Gy

### Dose rate:

for conventional 10<sup>-2</sup> Gy/s

for UHDR: 10<sup>2</sup>, 10<sup>4</sup>, 10<sup>6</sup>, 10<sup>8</sup> Gy/s

### PITZ time:

With triplicates 63 samples – 9 samples per hour – 7 h

Repeated three times – 7 h at daytime (app. 6:00 am – 6:00 pm)

# Challenges to solve

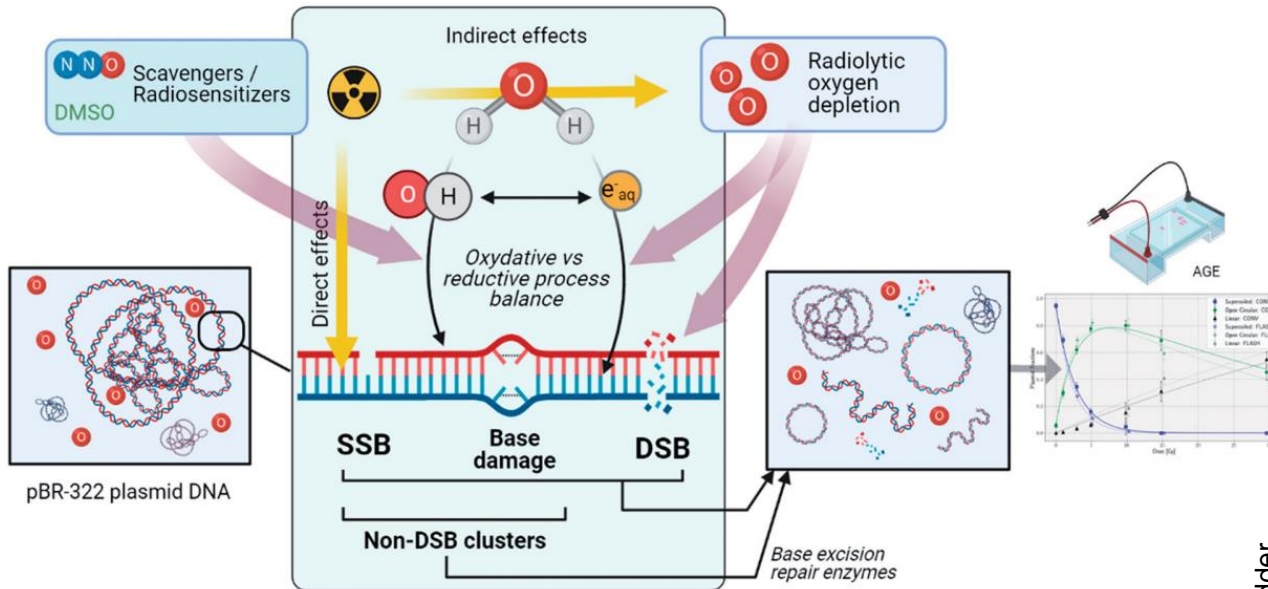
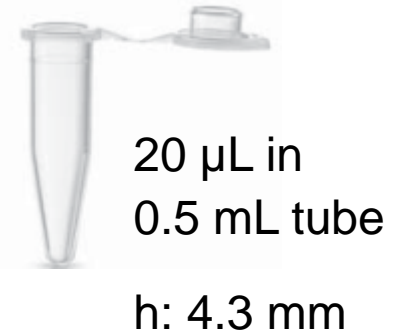
## In H<sub>2</sub>O<sub>2</sub> measurements:

? timing	Requirements	Plan
the reaction buffer (RB) should be added at the exact same time after irradiation for every sample	as fast as possible (200 s)	
after the RB addition, fluorescence measurements should be done at the exact same time point	30 min (up to 90 min)	<ul style="list-style-type: none"><li>bring reader to DESY? (which dates?)</li></ul>
no positive control	to test assay	<ul style="list-style-type: none"><li>make a calibration curve with stock H<sub>2</sub>O<sub>2</sub></li></ul>



# Biochemical effects of PITZ beam

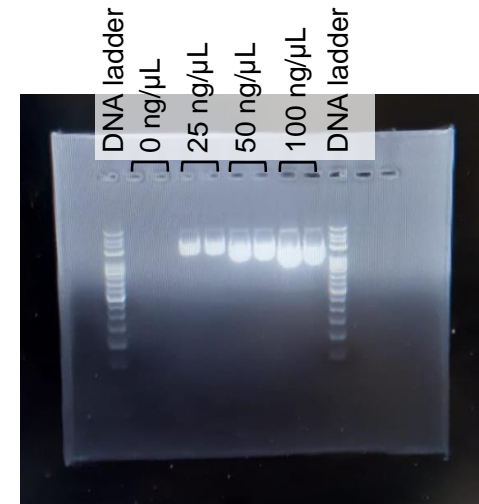
## DNA plasmid conformation



\* Alternatively the set-up can be adjusted to 0.2 mL tube

Kacem et. al. 2022 DOI: 10.1080/09553002.2021.2004328

- Gel electrophoresis



# Biochemical effects of PITZ beam

## DNA plasmid conformation



20  $\mu$ L in  
0.5 mL tube

h: 4.3 mm

### Dose:

0, 1, 2, 5, 10, 20, 30, 50 Gy

### Dose rate:

for conventional  $10^{-2}$  Gy/s

for UHDR:  $10^2$ ,  $10^4$ ,  $10^8$  Gy/s

### PITZ time:

With triplicates 66 samples – 10 samples per hour – 7 h

Repeated three times – 7 h at any time (samples should be stored at  $+4^{\circ}\text{C}$ )

# Challenges to solve

## In DNA plasmid study:

? cooling	Requirements	Plan
samples should be cooled for storage	4°C	
no positive control	to test assay	<ul style="list-style-type: none"><li>• test with UV light</li></ul>

# Biological effects of PITZ beam??

## Cell survival after irradiation

- Cancer cell lines:  
cervical carcinoma HeLa cells + lung adenocarcinoma A549 cells
- Normal cell lines:  
fetal kidney HEK293 cells + fetal lung fibroblast HEL299 cells

### Dose:

0, 2, 5, 10 Gy

### Dose rate:

for conventional  $10^{-2}$  Gy/s

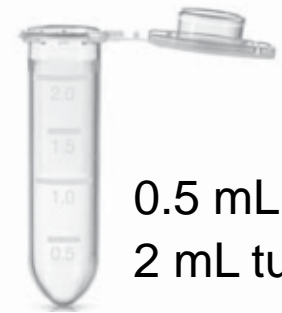
for UHDR:  $10^2$ ,  $10^8$  Gy/s

### PITZ time:

With triplicates 30 samples per cell line – 3 h per cell line

Repeated min three times – 3 h per cell line at daytime (app. 6:00 am – 6:00 pm)

- Colony formation – classic method, shows proliferation activity of cells
- Cell viability – high-throughput method, shows metabolic activity of cells



0.5 mL in  
2 mL tube

h: 11 mm

w: 10.5 mm

# Challenges to solve

## In cell-based study:

? timing	Requirements	Plan
cell should be prepared in a week		<ul style="list-style-type: none"><li>• I can prepapre cells for every day – 8-11 Nov.</li></ul>
cells should be held in tubes as short as possible	a few hours are ok	<ul style="list-style-type: none"><li>• In 1 h 20 min I need to know when to prepare them and bring to PITZ from TH Wildau,</li><li>• I will check longer times next week</li></ul>
colony formation assay	delivered reagents, 14 day-long analysis	<ul style="list-style-type: none"><li>• adapt the methodology</li><li>• check one (two) cell lines</li><li>• get extra hands</li></ul>

# Challenges to solve

? timing vs. manpower	Plan
<p>4-11 November – irradiation time + min 14 days for assays + min 10 days for analysis</p> <p>Deadline 28 November</p>	<ul style="list-style-type: none"><li>• a lot of work in parallel → extra hands</li><li>• <b>Yulia Komar</b>, Master student at Freie Uni / NTSU Kyiv – internship at TH Wildau / PITZ ?</li><li>• <b>Aleksandar Radivoievych</b>, PhD student at TH Wildau – support in cell culture</li></ul>

# Thank you for the attention!

